

Stress-Mediated Hepatotoxicity and the Protective Role of Phytomedicines

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ABSTRACT

Stress-mediated hepatotoxicity-liver injury driven or amplified by oxidative stress, endoplasmic reticulum (ER) stress, mitochondrial dysfunction, and inflammation-represents a convergent pathway for a broad range of chemical, biological and metabolic insults. Contributors include xenobiotics (drugs, environmental toxins), ischemia-reperfusion, viral infections, metabolic overload (nonalcoholic fatty liver disease), and lifestyle factors. The pathophysiology is characterized by redox imbalance, reactive oxygen and nitrogen species (ROS/RNS) formation, unfolded protein responses, impaired bioenergetics, and activation of innate immune signaling culminating in hepatocellular death (apoptosis, necrosis, necroptosis) and fibrogenic remodeling. Phytomedicines plant-derived extracts and purified phytochemicals offer multiple, often pleiotropic mechanisms for hepatoprotection: direct antioxidant activity, induction of endogenous cytoprotective pathways (Nrf2/ARE), attenuation of ER stress, preservation of mitochondrial function, anti-inflammatory effects (NF- κ B, inflammasome modulation), and inhibition of profibrogenic signaling. This review synthesizes mechanistic insights from preclinical models and the highest-quality clinical evidence to date, highlights leading phytochemical candidates (e.g., silymarin, curcumin, resveratrol, berberine, green tea catechins, quercetin, glycyrrhizin), discusses formulation and safety challenges, and proposes a research agenda to translate phytomedicines into evidence-based hepatoprotective interventions. We argue that, with standardized extracts, rigorous pharmacokinetic characterization and integrated safety monitoring, phytomedicines can complement conventional strategies to prevent or mitigate stress-mediated liver injury across clinical settings.

Keywords: hepatotoxicity, oxidative stress, phytomedicine, mitochondrial dysfunction, Nrf2

INTRODUCTION

The liver is uniquely exposed to chemical and metabolic stressors due to its central role in xenobiotic metabolism and nutrient handling. Diverse insults from prescription drugs and herbal products themselves, to ischemia, viral cytopathicity, alcohol, and lipotoxicity can trigger stress responses that overwhelm hepatocellular homeostatic mechanisms [1]. Central to many forms of liver injury is a disturbance of redox balance: excessive formation of reactive oxygen and nitrogen species (ROS/RNS) and failure of antioxidant defenses [2]. Concomitantly, ER stress and mitochondrial dysfunction create feed-forward loops that amplify cellular injury and activate inflammatory and fibrogenic cascades [3]. Collectively, these processes underlie drug-induced liver injury (DILI), progression of nonalcoholic steatohepatitis (NASH), and worse outcomes after ischemia-reperfusion or infection.

Phytomedicines the use of plant extracts or isolated phytochemicals for therapeutic benefit, have long historical roots and are gaining renewed scientific attention for liver protection [4]. Phytochemicals often exhibit multi-target activity that aligns conceptually with the multifactorial nature of stress-mediated hepatotoxicity: they scavenge ROS, induce phase II detoxification enzymes, stabilize membranes, modulate inflammatory signaling, and support mitochondrial resilience [5]. Yet translating these properties into consistent clinical benefit requires addressing heterogeneity in preparations, dose-ranging, pharmacokinetics, and safety. The following sections review the mechanistic basis of stress-mediated hepatotoxicity, summarize the hepatoprotective mechanisms attributed to key phytomedicines, evaluate the preclinical and clinical evidence, and outline practical considerations and research priorities.

2. Mechanisms of stress-mediated hepatotoxicity

Stress-mediated hepatotoxicity is not a single pathway but a network of interacting processes. The major mechanistic pillars include:

Oxidative and nitrosative stress. Phase I metabolism and pathological processes produce ROS/RNS that damage lipids, proteins, and nucleic acids [6]. Lipid peroxidation compromises membrane integrity and generates reactive aldehydes (e.g., 4-HNE) that further perturb signaling and mitochondrial function [7].

Endoplasmic reticulum (ER) stress and the unfolded protein response (UPR). Accumulation of misfolded proteins activates the UPR (IRE1 α , PERK, ATF6) [8]. While adaptive initially, prolonged UPR triggers CHOP-mediated apoptosis and amplifies oxidative stress [9].

Mitochondrial dysfunction and bioenergetic failure. Mitochondria are both sources and targets of ROS [3]. Loss of membrane potential, impaired electron transport, and opening of the mitochondrial permeability transition pore precipitate ATP depletion, release of proapoptotic factors (cytochrome c), and necrotic cell death [10].

Inflammation and innate immune activation. Damaged hepatocytes release damage-associated molecular patterns (DAMPs) that activate Kupffer cells and recruit neutrophils and monocytes [11]. Cytokines (TNF- α , IL-1 β) and inflammasome activation drive hepatocyte death and fibrogenesis [12].

Cell death modalities and fibrogenesis. Apoptosis, necrosis, necroptosis and pyroptosis occur depending on insult and context; chronic or massive parenchymal loss stimulates stellate cell activation and extracellular matrix deposition, leading to fibrosis [13]. Interconnectedness of these mechanisms means that interventions targeting one node can have broader protective effects—an important rationale for multi-target agents like phytochemicals.

3. Phytochemicals: mechanisms of hepatoprotection

Phytochemicals often exert hepatoprotective effects through several complementary mechanisms:

Antioxidant and phase II enzyme induction. Many flavonoids and polyphenols directly scavenge ROS and upregulate endogenous antioxidant defenses via nuclear factor erythroid 2-related factor 2 (Nrf2) [13]. Activation of the Nrf2-antioxidant response element (ARE) pathway increases expression of glutathione biosynthetic enzymes, glutathione S-transferases, heme oxygenase-1 (HO-1), and NAD(P)H:quinone oxidoreductase-1 (NQO1), restoring redox balance [14].

Mitigation of ER stress. Certain phytochemicals attenuate maladaptive UPR signaling—reducing CHOP expression and restoring ER-associated degradation—thereby lowering apoptosis triggered by persistent proteotoxic stress [15].

Mitochondrial protection. Compounds such as silymarin components, resveratrol, and certain alkaloids preserve mitochondrial membrane potential, enhance biogenesis (PGC-1 α pathways), and limit opening of the permeability transition pore, maintaining cellular ATP and preventing necrosis [17].

Anti-inflammatory and immunomodulatory effects. Suppression of NF- κ B signaling, inhibition of proinflammatory cytokine release, and modulation of inflammasome activity reduce immune-mediated amplification of hepatocyte injury [18].

Anti-fibrotic actions. By inhibiting stellate cell activation and matrix metalloproteinase/tissue inhibitor balance, some phytochemicals impede progression to fibrosis [19]. **Metabolic modulation.** In metabolic liver diseases, phytochemicals may improve insulin sensitivity, lipid handling, and fatty acid oxidation, addressing upstream drivers of lipotoxic stress [20]. These multi-modal actions make phytochemicals conceptually attractive for preventing or reducing stress-mediated hepatotoxicity across diverse clinical contexts.

4. Representative phytochemicals and evidence

Silymarin (milk thistle extract). Silymarin and its main flavonolignan, silybin, have long-standing use as hepatoprotectants [31]. Preclinical models show antioxidant, membrane-stabilizing and anti-fibrotic effects [21]; some clinical trials suggest improved liver enzyme profiles in chronic liver disease, though heterogeneity in extracts and endpoints limits firm conclusions [21].

Curcumin. The polyphenol curcumin exerts potent anti-inflammatory and antioxidant effects, modulates Nrf2 and NF- κ B pathways, and shows mitochondrial protective actions in animal models of toxin- and ischemia-induced liver injury [22]. Clinical data are promising in metabolic liver disease, but bioavailability challenges have spurred the development of improved formulations.

Resveratrol. A stilbene with mitochondrial and anti-inflammatory benefits, resveratrol enhances SIRT1/PGC-1 α signaling, supporting mitochondrial biogenesis and reducing oxidative injury in models of hepatic stress [23]. Human data are preliminary and dose selection remains complex.

Berberine. An isoquinoline alkaloid with metabolic and antioxidant effects, berberine improves lipid and glucose metabolism while attenuating toxin-induced hepatic injury in animals [24]; clinical evidence for liver protection is emerging but limited.

Green tea catechins (EGCG). Catechins show antioxidative and anti-inflammatory activity; however, concentrated extracts have been implicated in hepatotoxicity in rare cases, highlighting dose- and preparation-dependent safety considerations [25].

Quercetin, glycyrrhizin, saponins and other classes. Numerous other phytochemicals display hepatoprotective profiles in preclinical studies, ranging from antioxidant enzyme induction to anti-fibrotic signaling. Glycyrrhizin (licorice-derived) has a clinical history in viral hepatitis therapy in some regions [26].

Taken together, preclinical evidence robustly supports multiple phytochemicals as hepatoprotective in stress models, but consistent clinical proof, especially from large, well-controlled RCTs with standardized extracts and objective endpoints, remains limited.

5. Safety, standardization and formulation challenges

Key barriers to clinical translation include variable quality of botanical preparations, inconsistent phytochemical content, contamination/adulteration risks, and limited pharmacokinetic data. Some plant extracts cause idiosyncratic hepatotoxicity at high doses or with particular formulations (as seen with concentrated green tea extracts) [27]. Bioavailability is another major challenge: poorly absorbed compounds (e.g., curcumin) require formulation strategies (nanoparticles, phospholipid complexes) to reach therapeutic hepatic concentrations [28]. Drug–herb interactions via cytochrome P450 modulation or transporter effects are clinically relevant when phytomedicines are used alongside conventional drugs metabolized by the liver [29].

Regulatory heterogeneity complicates risk–benefit evaluation: in many jurisdictions, botanical products are marketed with limited premarket safety scrutiny. Standardization (certificate of analysis, batch testing), clinical-grade manufacturing (GMP) and active pharmacovigilance are prerequisites for responsible clinical use.

6. Translational pathways and research priorities

To harness phytomedicines against stress-mediated hepatotoxicity, the following priorities are critical: Standardized extracts and robust chemical characterization. Define active constituents and ensure batch-to-batch consistency. Pharmacokinetics and dose-finding. Determine hepatic exposure, metabolism, active metabolites and safe/effective dose ranges. Rigorous clinical trials with mechanistic endpoints. Trials should include biomarkers of oxidative stress (GSH/GSSG, 4-HNE), mitochondrial function, ER stress markers, and clinically meaningful outcomes (ALT/AST trajectories, histology, incidence of DILI, progression of fibrosis).

Safety surveillance and drug–herb interaction studies. Active reporting systems and prospective interaction studies will clarify risk profiles. Combination strategies and targeted populations. Phytomedicines may be most useful as adjuncts in early-stage NASH, perioperative ischemia–reperfusion protection, or as prophylaxis during known hepatotoxic drug exposure scenarios that lend themselves to focused clinical studies.

CONCLUSION

Stress-mediated hepatotoxicity is a mechanistic nexus underlying many forms of liver injury. Phytomedicines offer multi-targeted biologic actions that align with the complex pathophysiology of redox imbalance, ER stress, mitochondrial dysfunction and inflammation. Preclinical evidence is compelling; clinical translation requires standardized preparations, rigorous pharmacology, and well-designed trials incorporating mechanistic biomarkers and safety endpoints. With these steps, phytomedicines have the potential to complement conventional hepatoprotective strategies, offering accessible and biologically plausible tools to reduce liver injury in a variety of clinical contexts.

REFERENCES

1. Zhang Y, Qi Y, Huang S, Jiang X, Xiao W, Wang L, et al. Role of ER stress in Xenobiotic-Induced liver diseases and hepatotoxicity. *Oxidative Medicine and Cellular Longevity*. 2022;2022:1–11. doi:10.1155/2022/4640161
2. Uroko, Robert I., Adamude, Fatima A., Egba, Simeon I., Ani, Chijioke C and 1 Ekpenyong, James E. Hepatoprotective Effects of Methanol Extract of *Acanthus montanus* (acanthaceae) Leaves on Acetaminophen-Induced Liver Injury in Rats. *Pharmacologyonline*, 2020; 1: 248-260
3. Dery KJ, Chiu R, Kasargod A, Kupiec-Weglinski JW. Feedback loops shape oxidative and immune interactions in hepatic ischemia–reperfusion injury. *Antioxidants*. 2025;14(8):944. doi:10.3390/antiox14080944
4. Obasi, D.C., Abba, J.N., Anikete, U.C., Okoroh, P.N., Ugwu, O.P.C., Uti, D.E. (2025). Endogenous Plant signals and human Health: Molecular mechanisms, ecological functions, and therapeutic Prospects. *Biochemistry and Biophysics Reports*, 43(2025):102114. <https://doi.org/10.1016/j.bbrep.2025.102114>
5. Ochulor Okechukwu C., Njoku Obioma U., Uroko Robert I and Egba Simeon I. Nutritional composition of *Jatropha tanjorensis* leaves and effects of its aqueous extract on carbon tetrachloride induced oxidative stress in male Wistar albino rats. *Biomedical Research* 2018; 29(19): 3569-3576
6. Ugwu, CE., Sure, SM., Dike, CC., Okpoga, NA and Egba, SI. Phytochemical and *in vitro* antioxidant activities of methanol leave extract of *Alternanthera basiliana*. *Journal of Pharmacy Research*, 2018; 12(6): 835-839

7. Ayala A, Muñoz MF, Argüelles S. Lipid peroxidation: production, metabolism, and signaling mechanisms of malondialdehyde and 4-Hydroxy-2-Nonenal. *Oxidative Medicine and Cellular Longevity*. 2014; 2014:1–31. doi:10.1155/2014/360438
8. Sundaram A, Appathurai S, Plumb R, Mariappan M. Dynamic changes in complexes of IRE1 α , PERK, and ATF6 α during endoplasmic reticulum stress. *Molecular Biology of the Cell*. 2018;29(11):1376–88. doi:10.1091/mbc.e17-10-0594
9. Tabas I, Ron D. Integrating the mechanisms of apoptosis induced by endoplasmic reticulum stress. *Nature Cell Biology*. 2011;13(3):184–90. doi:10.1038/ncb0311-184
10. Giorgi C, Marchi S, Simoes ICM, Ren Z, Morciano G, Perrone M, et al. Mitochondria and reactive oxygen species in aging and Age-Related Diseases. *International Review of Cell and Molecular Biology*. 2018;209–344. doi:10.1016/bs.ircmb.2018.05.006
11. Woolbright BL, Jaeschke H. Mechanisms of inflammatory liver injury and Drug-Induced hepatotoxicity. *Current Pharmacology Reports*. 2018;4(5):346–57. doi:10.1007/s40495-018-0147-0
12. Taru V, Szabo G, Mehal W, Reiberger T. Inflammasomes in chronic liver disease: Hepatic injury, fibrosis progression and systemic inflammation. *Journal of Hepatology*. 2024;81(5):895–910. doi:10.1016/j.jhep.2024.06.016
13. Chakraborty JB, Oakley F, Walsh MJ. Mechanisms and biomarkers of apoptosis in liver disease and fibrosis. *International Journal of Hepatology*. 2012;2012:1–10. doi:10.1155/2012/648915
14. Jomova K, Alomar SY, Valko R, Liska J, Nepovimova E, Kuca K, et al. Flavonoids and their role in oxidative stress, inflammation, and human diseases. *Chemico-Biological Interactions*. 2025;111489. doi:10.1016/j.cbi.2025.111489
15. He F, Ru X, Wen T. NRF2, a transcription factor for stress response and beyond. *International Journal of Molecular Sciences*. 2020;21(13):4777. doi:10.3390/ijms21134777
16. Martucciello S, Masullo M, Cerulli A, Piacente S. Natural products targeting ER stress, and the functional link to mitochondria. *International Journal of Molecular Sciences*. 2020;21(6):1905. doi:10.3390/ijms21061905
17. Wu SK, Wang L, Wang F, Zhang J. Resveratrol improved mitochondrial biogenesis by activating SIRT1/PGC-1 α signal pathway in SAP. *Scientific Reports*. 2024;14(1). doi:10.1038/s41598-024-76825-9
18. Zhong Z, Umemura A, Sanchez-Lopez E, Liang S, Shalapour S, Wong J, et al. NF-KB restricts inflammasome activation via elimination of damaged mitochondria. *Cell*. 2016;164(5):896–910. doi:10.1016/j.cell.2015.12.057
19. Chan YT, Wang N, Tan HY, Li S, Feng Y. Targeting hepatic stellate cells for the treatment of liver fibrosis by natural products: Is it the dawning of a new era? *Frontiers in Pharmacology*. 2020;11. doi:10.3389/fphar.2020.00548
20. Tauil RB, Golono PT, De Lima EP, De Alvares Goulart R, Guiguer EL, Bechara MD, et al. Metabolic-Associated fatty liver disease: the influence of oxidative stress, inflammation, mitochondrial dysfunctions, and the role of polyphenols. *Pharmaceuticals*. 2024;17(10):1354. doi:10.3390/ph17101354
21. Jaffar HM, Al-Asmari F, Khan FA, Rahim MA, Zongo E. Silymarin: Unveiling its pharmacological spectrum and therapeutic potential in liver diseases—A comprehensive narrative review. *Food Science & Nutrition*. 2024;12(5):3097–111. doi:10.1002/fsn3.4010
22. Ghafouri-Fard S, Shoorei H, Bahroudi Z, Hussien BM, Talebi SF, Taheri M, et al. NRF2-Related therapeutic effects of curcumin in different disorders. *Biomolecules*. 2022;12(1):82. doi:10.3390/biom12010082
23. Chen Y, Zhang H, Ji S, Jia P, Chen Y, Li Y, et al. Resveratrol and its derivative pterostilbene attenuate oxidative stress-induced intestinal injury by improving mitochondrial redox homeostasis and function via SIRT1 signaling. *Free Radical Biology and Medicine*. 2021;177:1–14. doi:10.1016/j.freeradbiomed.2021.10.011
24. Hasanein P, Ghafari-Vahed M, Khodadadi I. Effects of isoquinoline alkaloid berberine on lipid peroxidation, antioxidant defense system, and liver damage induced by lead acetate in rats. *Redox Report*. 2016;22(1):42–50. doi:10.1080/13510002.2016.1140406
25. Ferrari E, Naponelli V. Catechins and Human Health: Breakthroughs from Clinical Trials. *Molecules*. 2025;30(15):3128. doi:10.3390/molecules30153128
26. Wahab S, Annadurai S, Abullais SS, Das G, Ahmad W, Ahmad MF, et al. Glycyrrhiza glabra (Licorice): A Comprehensive Review on Its Phytochemistry, Biological Activities, Clinical Evidence and Toxicology. *Plants*. 2021;10(12):2751. doi:10.3390/plants10122751
27. Mitaki, N.B., Fasogbon, I.V., Ojiakor, O.V., Makena, W., Ikuomola, E. O., Dangana, R.S., et al. (2025). A systematic review of plant-based therapy for the management of diabetes mellitus in the East Africa community. *Phytomedicine Plus*, 5(1): 100717. <https://doi.org/10.1016/j.phyplu.2024.100717>
28. Eyong ED, Iwara IA, Agwupuye EI, Agboola AR, Uti DE, Obio WA, et al. (2025) *In vitro* and *in silico* pharmaco-nutritional assessments of some lesser-known Nigerian nuts: *Persea americana*, *Tetracarpidium conophorum*, and *Terminalia catappa*. *PLoS ONE* 20(4): e0319756. <https://doi.org/10.1371/journal.pone.0319756>

29. Alum, E.U., Manjula, V.S., Uti, D.E., Echegu, D.A., Ugwu, O.P.C., Egba, S.I., Agu, P.C. (2025). Metabolomics-Driven Standardization of Herbal Medicine: Advances, Applications, and Sustainability Considerations. *Natural Product Communications*. 2025;20(8). doi:10.1177/1934578X251367650
30. Giorgi C, Marchi S, Simoes ICM, Ren Z, Morciano G, Perrone M, et al. Mitochondria and reactive oxygen species in aging and Age-Related Diseases. *International Review of Cell and Molecular Biology*. 2018;209–344. doi:10.1016/bs.ircmb.2018.05.006
31. Aja O. A., Egba S. I., Uhuo Emmanuel Nnaemeka, Alaebo Prince Ogocukwu, Mba Obinna Joseph, and Oriaku Chinwe Edith. Hepatoprotective potentials of aqueous chloroform and methanol leaf extracts *Whitfieldia lateritia* 2, 4-dinitrophenylhydrazine induced anaemia in rats. *Bio-research and Biotechnology*, 2022; 20(2) 1434–1445

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