

## Isolation of Seed-Borne Mycoflora from Cashew Nuts Vended in Selected Markets in Awka

Okigbo, R.N., \*Ezebo, R.O. and Obieze, I.V.

Department of Botany Faculty of Bioscience, Nnamdi Azikiwe University, P.M.B. 5025, Awka, Anambra State, Nigeria

\*Correspondence: [esau\\_056@yahoo.com](mailto:esau_056@yahoo.com); Phone No:+2348060044563

---

### ABSTRACT

This study investigated the mycoflora associated with Cashew nuts sold in Awka markets. Samples of Cashew nuts were purchased from local markets (Eke-Awka, Amaenyi and Good-Will). Samples were analysed for the moisture contents and the presence of fungi by adopting dilution plating method. Data were analysed using Analysis of Variance (ANOVA) via Statistical Analysis System (SAS) of Version 9.1. Means of treatment were compared using Duncan's Multiple Range Test (DMRT) at  $p < 0.05$ . The results of the study revealed that six fungal species were isolated which included *Fusarium oxysporum*, *Aspergillus niger*, *A. flavus*, *A. fumigatus*, *Penicillium oxalicum* and *Mucor* species. The results showed that Eke-Awka Market recorded the highest percentage moisture content of 6.8%, which was followed by Amaenyi Market with a percentage moisture content of 6.6%, while least was Goodwill Market with moisture content of 4.1%. The presence of fungi in stored food commodity deteriorates the nutritive value and secretes toxins injurious to human health. The occurrence of pathogenic fungi on cashew nuts could be avoided or diminished if proper farming, harvesting and storage methods were adopted.

**Keywords:** Isolation, Mycoflora, Cashew, Nut, Market, Awka

---

### INTRODUCTION

The cashew plant, *Anacardium occidentale* L., is a tropical evergreen, perennial tree with a darkish-green leathery foliage, crooked branches and very irregular crown, a small to medium sized tree belonging to the family Anacardiaceae [1]. For over 400 years after introduction, cashew trees were exploited mainly for apple and no commercial value was attached to the nuts [2]. The first commercial cashew planting in Nigeria was in the mid 1950 at Ogbe, Oji, Udi and Mbala by the defunct Eastern Nigeria Development Corporation (ENDC) and Iwo, Eruwa and Upper Ogun by the defunct Western Nigeria Development Corporation (WNDC) [3]. The cashew nut liquid as a by-product of processing cashew is mostly composed of anacardic acids [4] which have been used effectively against tooth abscesses due to their lethality to a wide range of gram-positive bacteria [5]. The nut oil from cashew seeds is used

topically as an anti-fungal agent, also for healing cracked heels as opined by [5]. [6], reported that consuming nuts at least four times a week resulted in a 37% reduced risk of coronary heart disease compared to those who never or seldom ate nuts. The cashew nut is a popular snack often eaten on its own and approximately 75% of their fat is unsaturated fatty acids which includes health [7]. In relation to the seed-borne mycoflora of cashew nut, fungi play a significant role in deteriorating the nutritive value of the nut by utilizing the nutrient for their growth. Plant based edible stuffs are very susceptible to fungal contamination [8]. Fungi are found in different food commodities including nuts and other parts of the plants and these results to economic losses on cashew nuts [9]. Several environmental factors like humidity and temperature during storage of these nuts influence the infestation by fungi and

aflatoxin production [10]. Contamination of foodstuffs by aflatoxins has created serious concerns on food safety in Africa and especially Nigeria [11]. Mycotoxins are diverse group of secondary metabolites produced by molds which contaminate foods and have toxic effects on the health of humans and animals. *Aspergillus* spp, *Penicillium* spp, *Rhizopus* spp and *Mucor* spp are the most frequent species recovered from non-disinfected cashew nuts [12]. Fungi grow to produce secondary metabolites under favorable chemical, physiological and environmental conditions, especially when temperature and moisture are suitable [13]; [14]; [15]. Mycotoxicoses are becoming increasingly implicated in human and animal pathology [16].

However, the mycotoxigenic *Aspergillus*, *Penicillium* and *Fusarium* are responsible for secretion of different metabolic toxic compounds [17]; [18] and could be considered the most serious fungal genera that contaminate cashew nuts [19]. [20], summarily stated that a proper study of mycoflora of cashew nuts may improve and establish a new variety of seeds as a necessary immediate measure to increase the average cashew nut yield in Nigeria. Considering the significance of cashew in the livelihood, the effect of mycoflora will cause a significant yield reduction of cashew in Nigeria, hence this study therefore, investigated the mycoflora associated with cashew nuts (*Anacardium occidentale* L.) vended in selected Markets in Awka.

## MATERIALS AND METHODS

### Sample Collection

Fresh Cashew fruits (*Anacardium occidentale* L.) samples were purchased from three different local markets in Awka axis namely; Eke-Awka Market, Amaenyi Market and Goodwill Market. The nuts were detached from the fruits and put in nylon bags separately and

brought into the laboratory of Department of Botany, Nnamdi Azikiwe University, Awka, where the nut shells were further opened to get the freshly enclosed edible cashew nuts for isolation of fungi.

### Preparation of Sample for Analysis

The fresh cashew nuts were washed with tap water and oven dried at a temperature of 60°C for 72 hours. The samples were ground into fine powder

using grinding machine after which the ground samples were sieved to obtain powdered processed sample used for the analysis.

### Determination of Moisture Content

The moisture content was determined according to AOAC (2000) method. An empty petri-dish was dried in an oven for 10 minutes and allowed to cool in a desiccator and then weighed ( $W_1$ ). Approximately 2g of the sample was weighed into the petri-dish ( $W_2$ ) and

placed in an oven at 105°C for 5 hours. It was then brought out, cooled in a desiccator continuously until a constant weighed is expressed ( $W_3$ ). The procedure was repeated for the different sample location of cashew nuts.

$$\% \text{ Moisture} = \frac{W_2 - W_3}{W_2 - W_1} \times 100$$

Where;

$W_1$  = weight of empty Petri-dish

$W_2$  = weight of sample + Petri-dish before drying

$W_3$  = weight of sample + Petri-dish after drying

### Mycological Analysis

Sample suspension was prepared by adding twenty grams of ground sample in 100 ml of sterile distilled water for two hours. Then, it was shaken for ten minutes using a mechanical shaker. Serial dilutions were prepared from  $10^{-1}$  to  $10^{-6}$  ml for each sample location under aseptic condition as fungal spores

sediment more quickly. One ml of appropriate dilution was transferred into petri dishes containing PDA medium using sterile pipette. Six replicates were produced. Then the plates were incubated at 27°C for 72 hours and then examined daily for the development of fungi growth.

### Sub-culturing/Identification of Test Fungi Pathogens

When growth has established, subcultures were prepared using inocula from different organisms in the mixed cultures to obtain a pure culture. This was done by transferring hyphal tips from the colony edge of the mixed cultures to fresh plates of PDA using flame sterilized blades. After sub-culturing, the plates were incubated at

27°C until pure cultures were obtained. The Petri dishes of pure cultures of the test fungi were then sealed with paraffin to prevent contamination. The resulting pure cultures were used for characterization and subsequent identification of the fungi isolates with the aid of a compound microscope and identification guides [21]; [22].

#### Fungal Count and Mean

The results of the fungal colonies were carefully counted and their means calculated. The number of colonies was

multiplied by the dilution factor, divided by the aliquot plated as follows:

$$\text{CFU/ML} = \frac{\text{Number of colonies} \times \text{the dilution factor}}{\text{Plating volume}}$$

#### Percentage Occurrence

Percentage occurrence was calculated using the following formula:

$$\% \text{ occurrence} = \frac{\text{No. of colonies of a particular fungi species in all plates}}{\text{Total no. of colonies of all the fungal colonies in all the plates}} \times 100$$

#### Statistical Analysis

Data were analysed using Analysis of Variance (ANOVA) via Statistical Analysis System (SAS) of Version 9.1. Means of

treatment were compared using Duncan's Multiple Range Test (DMRT) at  $p < 0.05$  [23].

### RESULTS

#### Isolation of Fungal Pathogens from Cashew Samples

The incidence of occurrence of fungi isolates associated with cashew nuts in Awka markets indicated that six fungi were isolated and they included

*Fusarium oxysporum*, *Aspergillus niger*, *A. flavus*, *A. fumigatus*, *Penicillium oxalicum* and *Mucor* species (Table 1).

**Table 1: Fungi isolated from cashew nuts from different locations in Awka**

S/N	Location	Fungal isolates
1	Eke-Awka Market	<i>Fusarium oxysporum</i> <i>A. niger</i> <i>A. flavus</i>
2	Amaenyi Market	<i>A. fumigatus</i> <i>A. niger</i> <i>A. flavus</i>
3	Goodwill Market	<i>A. niger</i> <i>Penicillium oxalicum</i> <i>Mucor</i> species

#### Fungal Count and Mean of the Cashew Nut Samples

The fungal colonies of the three different locations in Awka were observed and counted carefully. The data showed 133

fungal colonies from Eke-Awka, 90 fungal colonies from Amaenyi and 65 fungal colonies in Good-will market (Table 2).

**Table 2: Fungal colonies and total mean**

S/N	Location	Number of colonies	Mean colonies
1	Eke-Awka	133	1.33
2	Amaenyi	90	9.0
3	Goodwill	65	6.5

#### Percentage Occurrence

The results of the percentage occurrence of fungi isolates on cashew nuts obtained

from Eke-Awka Market revealed that *A.niger* showed highest occurrence of

45.1% while the least occurrence of 10.5% was recorded against *F. oxysporum* (Table 3). The results of the percentage occurrence of fungi isolates on cashew nuts obtained from Amaenyi Market showed that *A. niger* had highest occurrence of 43.3% while *A. fumigatus* showed the least occurrence of 16.7% (Table 4). The results of the percentage occurrence of fungi isolates on cashew nuts obtained from Goodwill Market

depicted that *Mucor* species showed highest occurrence of 53.8% while the least occurrence of 9.2% was recorded against *P. oxalicum* (Table 5). The results of the moisture contents of the cashew nuts obtained from three different locations showed that Eke-Awka Market recorded highest moisture content of 6.8% while the least moisture content of 4.1% was noticed against Goodwill Market (Table 6).

**Table 3: Percentage occurrence of fungi isolates on cashew nuts from Eke-Awka Market**

S/N	Fungi isolates	Individual occurrence	% occurrence
1	<i>Fusarium oxysporium</i>	14	10.5
2	<i>A. niger</i>	60	45.1
3	<i>A. flavus</i>	59	44.4
	Total colonies	133	

**Table 4: Percentage occurrence of fungi isolates on cashew nuts from Amaenyi Market**

S/N	Fungal isolates	Individual occurrence	% occurrence
1	<i>A. fumigatus</i>	15	16.7
2	<i>A. niger</i>	39	43.3
3	<i>A. flavus</i>	36	40.0
	Total colonies	90	

**Table 5: Percentage occurrence of fungi isolates on cashew nuts from Goodwill Market**

S/N	Fungal isolates	Individual occurrence	% occurrence
1	<i>A. niger</i>	24	37.0
2	<i>Penicillium oxalicum</i>	6	9.2
3	<i>Mucor</i> species	35	53.8
	Total colonies	65	

#### Moisture Content

**Table 6: Moisture contents of cashew nuts obtained from different locations**

S/N	Location	$W_1$ (g)	$W_2$	$W_3$	Moisture content %
1	Eke-Awka Market	6.192	7.786	7.676	6.8
2	Amaenyi Market	6.200	7.715	7.615	6.6
3	Goodwill Market	6.197	7.253	7.210	4.1

#### DISCUSSION

Cashew nut infection by pathogenic fungi has been reported in a number of studies and revealed a high risk due to contamination with mycotoxins [24]. It is a fact that plant based edible stuffs are very susceptible to fungal contamination [8]. From the results of this study, it was shown that six fungi were isolated which included *F. oxysporium*, *A. niger*, *A. flavus*, *A. fumigatus*, *P. oxysporium* and *Mucor* species. This finding is in line with reports of [25]. Most of the fungi were previously reported from cashew nuts in many parts of the world [26]; [24][27];

[25]. Incidence of these isolates depends on a number of factors including moisture content and storage time [26]. Low moisture levels limit mould growth after harvesting and during storage. The percentage moisture contents for the three sample locations revealed that Eke-Awka Market recorded highest moisture content of 6.8%, which was closely followed by Amaenyi Market with moisture content of 6.6%, while the least moisture content of 4.1% was noticed against Goodwill Market. This agrees with the preliminary study conducted by

some authors that moisture content below 5.8% is approximately equivalent to 70% relative humidity. This is generally considered the maximum level for safe storage [28]. However, samples from Eke-Awka and Amaenyi had moisture contents above the limit and consequently were predisposed to more fungal growth and mycotoxin contamination (Table 6). Cashew nut is one of the few commodities that travel a long distance between times of harvest and when consumed [29] and these can lead to the initiation of these fungal activities thereby causing losses of commercial and nutritional values in the nuts and most importantly endanger the

#### CONCLUSION AND RECOMMENDATION

This study demonstrated that cashew nuts vended in Awka are contaminated with several mycotoxigenic fungi which included *F. oxysporum*, *A. niger*, *A. flavus*, *A. fumigatus*, *P. oxalicum* and *Mucor* species. It is therefore, recommended that strict mycological

life of consumers by exposure to mycotoxins infestation. *Aspergillus* was represented by three species and showed the widest diversity among all isolated fungi which are *A. niger*, *A. flavus*, *A. fumigatus*, and followed by *Mucor* species, *Fusarium* and *Penicillium*. [30] stated that these fungal species are found common to the soil and different agricultural food commodities.

The contamination of toxigenic fungi and aflatoxins may occur from the soil during growth or harvesting or from the environment during storage and sale which are conducive for fungal growth proliferation and aflatoxin contamination [31].

hygiene measures should be established in cashew farming, harvesting and storage to minimize mycotoxin contamination and to discourage high humidity that favours the growth of these fungi.

#### REFERENCES

1. Cogley, L.S. and Steele, M. (1976). An Introduction to the Botany of Tropical Crops In 2<sup>nd</sup>ed, Longman Group Ltd., London. *Mycology and Spoilage of Retail Cashew Nuts*. Pp. 369-373.
2. Aliyu, O.M. (2012). Genetic Diversity of Nigeria Cashew Germplasm. *Academic Journals*, **10**(50): 196-209.
3. Asogwa, E.U., Anikwe, J.C., Ndubuaku, C.N. and Okelana, F.A. (2009). Distribution and damage characteristics of an emerging insect pest of cashew, *Plocaederus ferrugineus* L. (Coleoptera: Cerambycidae) in Nigeria: A preliminary report. *African Journal of Biotechnology*, **8**(1):053-058.
4. Alexander, T.H. (2008). A Nutty Chemical. *SM Journal of Biology*, **86**:26-27.  
Association of Official Analytical Chemists (2000). *Official Method of Analysis of Association of Official Analytical Chemists*. 17<sup>th</sup> ed., AOAC International. Gaithersburg. 2200p.
5. Akash, P.D. Vishal, D. and Joshi, A.B. (2009). Antimicrobial screening of different extracts of *Anacardium occidentale* Linn. Leaves. *International Journal of Chemistry and Technological Research*, **1**:856-858.
6. Kelly, J.H. and Sabate, J. (2006). Nuts and coronary heart disease perspective. *Epidemiological Journal*, **2**:61-67.
7. Bes-Rastrollo, M., Sabate, J., Gomez, G.E., Alonso, A., Martinez, J.A. and Martinez-Gonzalez, M.A. (2007). Nut consumption and weight gain in a Mediterranean cohort. *The Sun Study Obesity*, **1**:107-116.
8. Raghu, A. and Eadie, T. (1978). Evaluation of potential for aflatoxin occurrence on celery, cauliflower, lettuce and taro root inoculated with *Aspergillus flavus* and *A. parasiticus*. *Journal - Association of Official Analytical Chemists*, **61**(4):998-1001.
9. Pitt, J.I. and Hocking, A.D. (2009). Fungi and food spoilage. *African Journal*, **5**:9-111.
10. Drusch and Ragab (2003). Mycotoxins in fruits, fruit juices and dried fruit. *Journal of Food Protection*, **66**:1514-1527.

11. Okigbo, R.N. (2002). Mycoflora of tuber surface of white yam (*Dioscorea rotundata*) and post-harvest control of pathogens with *Bacillus subtilis*. *African Journal of Biotechnology*, 4(8): 804-807.
12. Adebayo, L.O. and Diyaolu, S.A. (2003). Mycology and spoilage of retail cashew nuts. *African Journal Biotechnology*, 2(10):369-37.
13. Afsah-Hejri, L., Jinap, S., Hajeb, P., Radu, S. and Shakibazadeh, S.H. (2013). A Review on Mycotoxins in Food and Feed. *Biological Sciences and Pharmaceutical Research*, 8(2):28-36.
14. Ukwuru, M.U., Ohaegbu, C.G. and Muritala, A. (2017). An Overview of Mycotoxin Contamination of Foods and Feeds. *Journal of Biochemistry and Microbial Toxicology*, 1(1):1-11.
15. Leslie, J.F. (2005). Mycotoxin contamination in foods in West and Central Africa. *Journal of Microbiology Biotechnology*, 24(3): 297-312.
16. Bacha, H., Hadidane, R., Creppy, E.E., Regnault, C., Ellouze, F. and Dirheimer, G. (1988). Monitoring and identification of fungal toxins in food products, animal feed and cereals in Tunisia. *Journal of Stored Products Research*, 24:199-206.
17. El-Samawaty, M.A., Yassin, M.A., Bahkali, A., Moslem, M.A. and Abd-Elsalam, K.A. (2011). Biofungal activity of *Aloe vera* sap against mycotoxigenic seed-borne fungi. *Fresenius Environmental Bulletin*, 20(6):1352-1359.
18. Yassin, M.A., El-Samawaty, A.M., Bahkali, A., Moslem, M., Abd-Elsalam, K.A. and Hyde, K.D. (2010). Mycotoxin-producing fungi occurring in sorghum grains from Saudi Arabia. *Fungal Diversity*, 44:45-52.
19. Suleiman, M.N. (2010). Occurrence and distribution of fungi associated with bio-deterioration of cashew in the eastern senatorial district, Kogi State, Nigeria. *Applied Science Research*, 2(5):462-465.
20. Desci, A.R. (2008). *Molecular diversity and phenotype of selected cashew genotypes of Goa, and physiological response, in situ moisture conservation*. College of Agriculture, Dharwad University of Agricultural Sciences, Dharwad. P. 580.
21. Sulton, B.C. (1980). *The Coelomycetes Fungi Imperfecti with Pycnidia acervuli and Stromata*. Common Wealth Mycological Institute, Kew, Surrey, England. 696p.
22. Barnett, H.L. and Hunter, B.B. (1987). *Illustrated General of Imperfect Fungi* (4<sup>th</sup> Edition). Burgess Publishing Company. 218p.
23. Duncan, D. B. (1955). Multiple Range and Multiple F-test. *Biometrics*, 11: 1-42.
24. Mohammed, S.A. (2012). Evaluation of fungal flora and mycotoxin potential associated with postharvest handlings of cashew nut. *Journal of Science*, 8(12): 525-534.
25. Adeniyi, D.O. and Adediji, A.R. (2015). Evaluation of fungal flora and mycotoxin potential associated with postharvest handlings of cashew nut. *Scholars Research Library*, 2:30- 33.
26. Chelack, W.S., Borsa, J., Marquardt, R. and Frohlich, A.A. (1991). Evaluation of fungal flora and mycotoxin potential associated with postharvest handlings of cashew nut. *Applied and Environmental Microbiology*, 57:2492-2496.
27. Yilmaz, I. and Aluc, M. (2014). Determination of Aflatoxin levels in cashews on Turkish markets. *Food Journal*, 7:321-323.
28. Pixton, S.W. (1982). The importance of moisture and equilibrium relative humidity in stored products. *African Journal of Biotechnology*, 2(10): 369-373.
29. Pillao, P.B. (2008). *Cashew Handbook on Evaluation of fungal flora and mycotoxin potential associated with postharvest handlings of Cashew Nut*. Foretell Business Solutions Private Limited, Bangalore. 33p.

30. Srivastava, M., Pande, S., Srivastava, L. and Srivastava, C. (2014). Fungal infestation in some dry fruits during storage in different seasons. *International Journal of Multidisciplinary and Current research*, 14:23-27.

31. Reddy, Rabelo, M.C. and Fontes, M.L. (2011). Evaluation of fungal flora and mycotoxin potential associated with postharvest handling of cashew nut. *Journal of Plant Physiology*, 19(4):3-6.

**Okigbo, R.N., Ezebo, R.O. and Obieze, I.V. (2023). Isolation of Seed-Borne Mycoflora from Cashew Nuts Vended in Selected Markets in Awka. IDOSR JOURNAL OF APPLIED SCIENCES 8(3) 140-146. <https://doi.org/10.59298/IDOSR/2023/10.2.1420>**