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Synthesis, Characterization and anti-Microbial Studies of Metal (II) Complexes with (E)-4-methoxy-N-(4-methoxybenzylidene) aniline

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ABSTRACT

Some divalent transition metal complexes was synthesized by the reaction of the chlorides of Co(II), Fe(II), Mn(II), and Ni(II) with Schiff base derived from the reaction of 4methoxybenzaldehyde with 4-methoxyaniline in methanolic solution. The Schiff base, L was obtained as grey crystals with melting point of 153°C. The metal(II) complexes were all coloured, ML melted in the range of 171-177°C. A conductivity measurement for the complexes in DMSO was in the range of 77.58-85.10 Ω⁻¹mo¹⁻¹cm². Thelarge values for L suggest the presence of anion outside the coordination sphere. All the complexes were found to be paramagnetic as observed from the values of their magnetic moments and also soluble in most common solvents. The infra-red spectral analysis indicates complexation has taken place. The metal-ligand ratio in all the complexes was 1:2 from Job's method of continuous variation results. The percentage composition of the metals and their corresponding ligands were found from the complexes gravimetrically. The results obtained were in good agreement with the calculated values. The ligand and its corresponding complexes were screened in-vitro for antibacterial activity against some pathogenic bacteria: Escherichia coli, Aspergillus fumigatus, Salmonella typhi, Staphylococcus aureus, Aspergillus flavus and Mucor species using the paper-discs diffusion method. Inhibition properties of the complexes showed that the complexes have higher antimicrobial activity than the ligand which could be attributed to chelation.

Keywords: 4-methoxybenzaldehyde, 4-methoxyaniline, Schiff bases, antimicrobial activity

INTRODUCTION

Schiff bases are an important class of ligands derived from amino and carbonyl compounds that coordinate to metal ions via azomethine nitrogen and have been studied extensively. The C=N linkage is

essential for biological activity, several azomethines were reported to possess remarkable antibacterial, antifungal, anticancer and diuretic activities [1].

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Schiff bases have wide applications in food industry, dye industry, analytical chemistry, catalysis, fungicidal, agrochemical and biological activities [2, 3]. Schiff-base complexes are considered to be among the most important stereochemical models in main group and transition metal coordination chemistry due to their preparative accessibility and structural variety [4].

Para-anisidine (*p*-anisidine) 4methoxyaniline, a grey-brown solid, is the most toxic of the three isomers of anisidine and causes blood damage upon oral ingestion, inhalation or skin contact [5]. *P*-anisaldehyde 4-methoxy or benzaldehyde is an organic compound (slightly yellow oily liquid) that occurs naturally in the fennel and anise plants. Herein is reported the synthesis, characterization and anti-microbial studies of metal (II) complexes with (E)-4methoxy-N-(4-methoxybenzylidene) aniline derived from 4-methoxyaniline and 4-methoxybenzaldehyde

Materials and Methods

All chemicals/reagents the used throughout this work were of analar They includes; 4grade. methoxybenzaldehyde, 4-methoxyaniline, ethanol, DMSO, $P_{3}O_{5}$ (phosphorus pentaoxide), methanol. The materials used includes, hot plate with magnetic stirrer, condenser and round bottom flask, beakers, conical flasks, funnel, filter papers.

Synthesis of the Schiff base ligand

The Schiff base was synthesized as previously described [6, 7, 8] with a slight modification. The Schiff base was synthesized by the condensation of 4methoxybenzaldehyde (6mmol, 0.8169g) with 4-methoxyaniline (6mmol, 0.7389g), in a 30cm3 methanolic solution (1:1) molar ratio to give L. The mixture was refluxed for one hour after which it was cooled to room temperature. The product formed was filtered, washed with 3x5ml portions of ethanol and dried over anhydrous phosphorus pentaoxide in a desiccator.

Synthesis of the M (II) complexes

The complexes were synthesized by reacting with Metal (II) salt and the ligand in a 1:2 molar ratio. To a solution of the Schiff base ligands, L in 10ml methanol, 3mmol of CoCl2.6H2O, FeCl2.4H2O, MnCl2.4H2O or NiCl2.6H2O in 25ml methanol was added with constant stirring. The mixture was refluxed for 2 hours using a hot plate with magnetic stirrer, after which it was allowed to cool at room temperature and the product formed was filtered, washed with 3x5ml portions of methanol and stored in a desiccator containing anhydrous phosphorus pentaoxide.

Gravimetric determination of metals in the complex compounds

The samples were separately digested as described by [8] by weighing 0.2g of the desired metal complex into a 100cm³ beaker containing 25cm³ of distilled water and 5cm³ of concentrated nitric acid (HNO₃, specific gravity 1.42g/cm³). The mixture was heated to about dryness and the contents of the beaker were allowed to cool to room temperature after which

25cm³ of distilled water was added. The mixture was stirred, filtered and the filtrate collected and used for their determinations.

Determination of Manganese as Mn(NH₄)PO₄.H₂O

Distilled water was added to the filtrate obtained from the digested manganese (II) complex compound and made up to the mark in a 100cm3 volumetric flask with more distilled water. The solution was transferred to a 250cm3 conical flask and dilute ammonia solution was added to neutralize the filtrate after which 10g of ammonium chloride, excess diammonium hydrogen phosphate (NH₄)₂HPO₄ and some few drops of (1:3) hydrochloric acid were added respectively. The mixture was heated to 95°C followed by drop wise addition of dilute aqueous ammonia solution with constant stirring until a precipitate of [Mn₂(PO₄)₂] began to form. The addition of ammonia solution was stopped immediately while heating and stirring continued in order to ensure that the precipitation of MnNH, PO, .H, O. The beaker and its contents were allowed to cool to room temperature. The precipitate

was filtered, washed with 1% ammonium nitrate solution and the product heated to a constant weight. It was weighed as MnNH,PO,.H,O, [8].

Determination of Iron as Fe₂O₃

Iron was determined by adding ammonia solution (6M) to the hot solution of the complex with constant stirringuntil the solution was slightly basic (this was indicated by a pH probe). The probe was rinsed back into the beaker before removing it so as to not lose any iron. The precipitate was digested by boiling for 5 minutes (by flocculation) and allowed to settle. The solution was stirred and filtered. The precipitate was washed with hot 1% NH₄NO₃. A small volume of the filtrate was transferred into a beaker and few drops of 0.1M AgNO₃ added to test for completion of precipitation which was indicated by non-formation of a white precipitate. The precipitate was filtered using a filter paper which was folded and burnt in a crucible at red heat for 15minutes to obtain the Fe,O, residue. Once the sample has been ignited, the

crucible was cooled completely before its mass was accurately measured [9].

Determination of Cobalt as Co[Hg(SCN),]

4.8ml of mercury (II) chloride solution was added to the solution of the complex with constant stirring followed by addition of 5.5cm³ammonium thiocyanate solution for 1 - 3 minutes, a dark-blue precipitate was formed. Stirring was continued for a further 3 minutes before the solution was allowed to stand for2 hours at room temperature. The precipitates formed were collected in aweighed sintered glass (porosity no.4), washed with 3cm3 of a dilute solutionof ammonium thiocyanate and finally with 5cm3 ice-cold water. It was then dried at 100C and weighed as Co[Hg(SCN),] [8].

Determination of Nickel as Ni (HDMG)₂

0.35g of pure ammonium nickel sulphate (NH₄)₂SO₄.NiSO₄.6H₂O was weighed and dissolved in 25cm³ water contained in 500cm³ beaker. 5cm³ of HCl (1:1) was added and then diluted to 200cm³with more distilled water before heated to 80°C, a slight excess

ofdimethylglyoximate (DMG) solution was added to the solution immediately followed by drop wise addition of dilute ammonia solution with constant stirring until precipitation took place. The mixture was allowed to stand on a steam bath for 30 minutes for the red DMGnickel complex to settle down. The precipitate was allowed to stand for another 60 minutes to cool to the room temperature. The cold solution was filtered through a sintered-glass 120°C. previously heated to The precipitates was washed with cold water until free from chloride ions and dried at 110°C for 50 minutes. It was then allowed to cool in a desiccator and reweighed until a constant weight was achieved. It wasweighed finally asNi(C₄H₂O₂N₂)₂ [8].

Determination of Chloride as [AgCI]

The percentage of chlorine in the sample was determined by weighing about 0.2g of the complex into a 250cm³beaker with astirring rod and covered with a watch glass. Then about 150 cm³ of water and 0.5cm³ of concentrated nitric acid were added to the complex and stirred until the

solids dissolved. 0.1M of silver nitrate solution was added slowly to the cold solution of the complex with constant stirring. The determination was done in a dark environment. The mixture was heated to nearly boiling temperature with constant stirring until the precipitate formed coagulated and the supernatant liquid was clear (between 2-3 minutes). Few drops of 0.1M silver nitrate solution were added to the supernatant as a confirmatory test to check for complete of precipitation (absence further precipitates confirmed the end of the test). The beaker with its content was kept in the dark and allowed to cool for 1 hour before filtration. A sintered glass filtering crucible (no. 4) was washed, dried (at 130°C) and cooled in a desiccator was weighed separately. The precipitates were washed 3 times with about 30cm³ of cold and very dilute nitric acid (0.5cm³ of the concentrated acid was dissolved in200cm³ of water) and collected in the weighed crucible. The precipitate was washed again in the crucible with cold and very dilute nitric acid (0.05M) until 5cm3 of the

washings collected in a test tube gave no turbidity when two drops of 0.1M hydrochloric acid was added to it. The crucible with its contents were placed in an oven and dried at 140°C for 60 minutes before it was finally cooled in a desiccator and weighed as AgCl. The heating and cooling was repeated until a constant weight was attained. The percentage of chlorine in the sample was then calculated [9, 10].

Antibacterial Activity

The ligand and its complexes were dissolved separately in DMSO to have five different concentrations (Table 7a-b). They were placed on the surface of the culture and incubated at 37°C for 24hrs; the diameter of the zone of inhibition surrounding the disc was taken as the measure of inhibitory power of the compound against a particular test organism.

RESULTS AND DISCUSSION

Table 1: Physical properties of the ligand and its complexes

Compour	nds Formular	Formular weig	ht (g) Colour Yield (%)	M.pt (°C)
L	C ₁₅ H ₁₅ NO ₂	241.28	Grey 84.64	153
CoL	[CoL ₂]	541.50	Yellow 84.94	171
FeL	[FeL ₂]	538.41	Brown 83.42	176
MnL	[MnL ₂]	537.51	Yellow 90.87	174
NiL	$[NiL_2]$	541.26	Green 89.2	0 177

Key: $L = C_{15}H_{15}NO_2$

Table 2: Infrared Spectral and Gravimetric data of theligand and their complexes

Compound	v C=N v M-N v M-O v O-CH ₃ % metal in the complexes										
				Calcul	lated Observ	ved					
L	1623			2942							
CoL	1610	645	536	2939	10.88	10.96					
FeL	1620	644	533	2943	10.37	10.44					
MnL	1623	632	537	2940	10.22	10.30					
NiL	1620	637	537	2943	10.84	10.93					
Key:	$L = C_{15}H$	I ₁₅ NO ₂									

Table 3: Molar conductivity of the Schiff base complexes

Compound	Concentration, C	Specific conductance (S)	Molar conductance
	(mol)		$(\Omega^{-1} mol^{-1} cm^2)$
CoL	1 x 10 ⁻³	2.58 x 10 ⁻⁶	2.58
FeL	1 x 10 ⁻³	2.10×10^{-6}	2.10
MnL	1 x 10 ⁻³	1.95 x 10 ⁻⁶	1.95
NiL	1 x 10 ⁻³	1.55 x 10 ⁻⁶	1.55

Key: $L = C_{15}H_{15}NO_2$

Table 4: Magnetic Susceptibility of the complexes

Compounds Gram susceptibility, Xg Molar susceptibility, Xm Effective magnetic									
erg c ⁻² g ⁻¹		moment (µeff) B.M							
CoL	1.62 x 10 ⁻⁵	8.75×10^{-3}	4.62						
FeL	2.04 x 10 ⁻⁵	1.10 x 10 ⁻²	5.17						
MnL	1.67 x 10 ⁻⁵	8.96 x 10 ⁻³	4.68						
NiL	2.18 x 10 ⁻⁵	1.20 x 10 ⁻²	5.34						
Key:	$L = C_{15}H_{15}NO_2$								

Table 5a: Job's method of continuous variation results of the metal(II) complexes

Co(II) Complex	Fe(II) Complex	Mn(II) Complex	Ni(II) Complex							
$(\lambda max = 681.63nm) (\lambda max = 421.24nm) (\lambda max = 900.60nm) (\lambda max = 420.87nm)$										
XL Abs	XL Abs	XL Abs	XL Abs							
0.0830 0.1174	0.0830 0.1521	0.0830 0.0899	0.0830 0.1790							
0.1670 0.2243	0.1670 0.1722	0.1670 0.1034	0.1670 0.2483							
0.2500 0.2798	0.2500 0.1828	0.2500 0.1202	0.2500 0.2648							
0.3330 0.3394	0.3330 0.1998	0.3330 0.1364	0.3330 0.3201							
0.4170 0.3906	0.4170 0.2201	0.4170 0.1497	0.4170 0.3506							
0.5000 0.4425	0.5000 0.2446	0.5000 0.1672	0.5000 0.4011							
X _L = Mole fraction;	X _L = Mole fraction; Abs = Absorbance									

Table 5b (cont): Job's method of continuous variation results of the metal (II) complexes

Co(II) Complex	Fe(II) Complex	Mn(II) Complex	Ni(II) Complex						
$(\lambda max = 681.63nm) (\lambda max = 421.24nm) (\lambda max = 900.60nm) (\lambda max = 420.87nm)$									
XL Abs	XL Abs	XL Abs	XL Abs						
0.5830 0.5021	0.5830 0.2622	0.5830 0.1882	0.5830 0.4535						
0.6670 0.5716	0.6670 0.2898	0.6670 0.1984	0.6670 0.4901						
0.7500 0.4515	0.7500 0.2481	0.7500 0.1721	0.7500 0.3255						
0.8330 0.2719	0.8330 0.1921	0.8330 0.1501	0.8330 0.2309						
0.9170 0.1009	0.9170 0.1525	0.9170 0.1172	0.9170 0.1089						
XL = Mole fraction;	; Abs = Absorbance								

Table 6: Solubility of the ligand/complexes in water and some common organic solvents

Solvents	L	CoL	FeL	MnL	NiL
Acetone	S	S	S	S	S
CCl4	S	IS	SS	SS	SS
Chloroform	S	S	S	S	S
Diethylether	IS	S	S	S	S
DMF	S	\mathbf{S}	SS	S	S
DMSO	S	S	S	S	S
Ethanol	IS	IS	IS	IS	IS
Formaldehyde	IS	IS	SS	IS	IS
Acetic acid	S	\mathbf{S}	S	S	S
Methanol	IS	IS	IS	IS	IS
Pet. Ether	IS	IS	IS	SS	IS
Water	IS	IS	IS	IS	IS

Key: $L = C_{15}H_{15}NO_2$ IS = Insoluble S = Soluble SS = slightly soluble

Table 7a: Antibacterial activity of the ligands/complexes

Compound	Organism	Concentration (μg)/Zone of inhibition (mm)						
		5000	400	3000	2000	1000	Control (mm)	
L	S. aureus	11	09	07	NZI	NZI	28 (Ampiclox)	
L	E. coli	08	07	04	NZI	NZI	31 (Ampiclox)	
L	S. typhi NZI	NZI	NZI	NZI	NZI	34 (An	mpiclox)	
L	A. fumigatus	NZI	NZI	NZI	NZI	NZI	28 (Ketoconazole)	
L	A. flavus	NZI	NZI	NZI	NZI	NZI	25 (Ketoconazole)	
L	Mucor. s	NZI	NZI	NZI	NZI	NZI	33 (Ketoconazole)	
CoL	S. aureus	11	09	07	NZI	NZI	28	
CoL	E. coli	10	09	06	NZI	NZI	31	
CoL	S. typhi 10	08	07	05	NZI	34		
CoL	A. fumigatus	08	06	05	03	NZI	28 (Ketoconazole)	
CoL	A. flavus	NZI	NZI	NZI	NZI	NZI	25 (Ketoconazole)	
CoL	Mucor. s	NZI	NZI	NZI	NZI	NZI	33 (Ketoconazole)	
FeL	S. aureus	09	07	05	NZI	NZI	28(Ampiclox)	
FeL	E. coli	10	08	07	NZI	NZI	31(Ampiclox)	
FeL	S. typhi 11	10	07	NZI	NZI	34(Am	npiclox)	
FeL	A. fumigatus	NZI	NZI	NZI	NZI	NZI	28 (Ketoconazole)	
FeL	A. flavus	NZI	NZI	NZI	NZI	NZI	25 (Ketoconazole)	
FeL	Mucor. s	NZI	NZI	NZI	NZI	NZI	33 (Ketoconazole)	
MnL	S. aureus	NZI	NZI	NZI	NZI	NZI	28(Ampiclox)	
MnL	E. coli	NZI	NZI	NZI	NZI	NZI	31(Ampiclox)	
MnL	S. typhi NZI	NZI	NZI	NZI	NZI	34(Am	npiclox)	
MnL	A. fumigatus	NZI	NZI	NZI	NZI	NZI	28 (Ketoconazole)	
MnL	A. flavus	NZI	NZI	NZI	NZI	NZI	25 (Ketoconazole)	
MnL	Mucor. s	NZI	NZI	NZI	NZI	NZI	33 (Ketoconazole)	
NiL	S. aureus	12	10	07	NZI	NZI	28 (Ampiclox)	
NiL	E. coli	10	08	07	NZI	NZI	31 (Ampiclox)	
Key: $L = C_1$	$_{15}H_{15}NO_2$ NZI =	No zone	of inhib	ition	Ampic	elox = (50)	Omg per disc)	
Ketoconazole = (200mg per disc)								

Table 7b (cont): Antibacterial activity of the ligands/complexes								
Compound	Organism	Conce	Concentration (µg)/Zone of inhibition (mm)					
		5000	400	3000	2000	1000	Control (mm)	
NiL	S. typhi	10	08	07	NZI	NZI	34 (Ampiclox)	
NiL	A. fumigatus	09	07	03	NZI	NZI	28 (Ketoconazole)	
NiL	A. flavus	NZI	NZI	NZI	NZI	NZI	25 (Ketoconazole)	
NiL	Mucor. S	NZI	NZI	NZI	NZI	NZI	33 (Ketoconazole)	
Key: $L = C_{15}H_{15}NO_2$ $NZI = No$ zone of inhibition Ampiclox = (500mg per disc)								
Ketoconazole = (200mg per disc)								

Schiff base, L

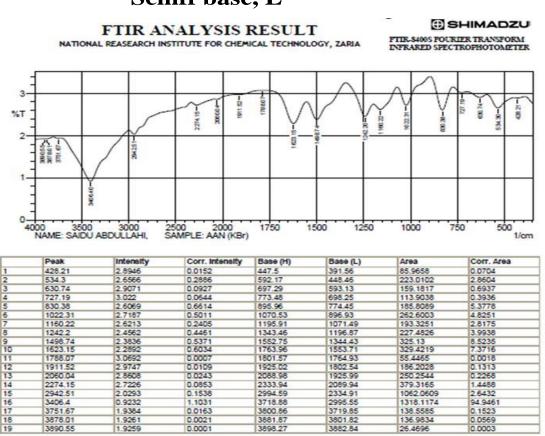


Fig 1: FTIR result for L

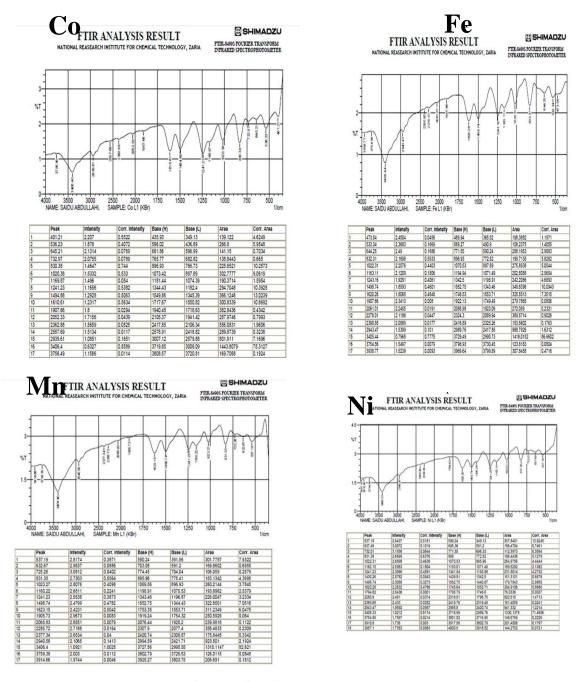


Fig 2: FTIR results for the Schiff base metal complexes

DISCUSSIONS

The interaction between 4methoxybenzaldehyde with p-anisidine gave a grey coloured crystals of Schiff base L, called (E)-4-methoxy-N-(4methoxybenzylidene) aniline. The percentage yield of the Schiff base ligand was 84.64% and with a melting point of 153-174°C. The reactions of the Schiff base L. with the respective metal (II) chlorides (NiCl₂.6H₂O, CoCl₂.6H₂O, FeCl₂.4H₂O or MnCl₂.4H₂O) produced complexes that are variously coloured with percentage yield in the range 83-90% . The ligand and its complexes have sharp melting points, an indication of purity for the substances prepared. The complexes are all anhydrous and air stable. The Schiff base and their complexes were found to be soluble in most solvents (Table 6). The molar conductance (Table 3) of complexes the in dimethylsulphoxide was in the range of 77.58-85.10 Ω^{-1} mol⁻¹cm² for L, which suggests the presence of anion outside the coordination sphere. These values are in good agreement with those reported

earlier [11,12, 13, 14, 15]. The magnetic moment values of the complexes obtained (Table 4) was in the range of 4.62-5.34 B.M and was an indication of paramagnetic nature of the complexes due to the presence of unpaired electrons in the partially filled d-orbitals. The X value confirmed positive that substances have unpaired electrons; this is in agreement with what has been reported earlier [16]. Gravimetric analysis data (Table 2) for the complexes obtained showed that the complexes analyzed as ML2 and the results obtained were in good agreement with the calculated values. A plot of absorbance (y axis) versus the mole fraction (x axis) of the ligand revealed a stoichiometric values of 1:2 metal-ligand ratio.

Infrared: Table 2 revealed the infrared spectra of the ligand which showed a characteristic band at 1623-1627 cm⁻¹ which is attributable to υ C=N band. This band shifted to a lower range of frequencies 1610-1623cm⁻¹ in the complexes ML; this indicates involvement

of theazomethine nitrogen in bonding, the result bands are similar to the values reported by [17, 18]. The band at 2942cm ¹ in the ligand, which was present in the spectra of its corresponding complexes was assigned to υ O-CH and it indicated the non-involvement of the methyl group in complexation as also reported by [19]. The coordination through nitrogen of azomethine and oxygen of υ O-CH₂ group of the ligand with its complexes are further evidenced by the appearance in the complexes of new but low frequency non-ligand bands around 632-645cm⁻¹ and 533-537cm⁻¹ assigned to υ M-N and υ M-O respectively. These bands were absent in the ligand. The shifting of the band at

1623 cm⁻¹ to lower values in all the complexes, low frequency bands (around 632-645cm⁻¹) and the vibration centered around 2942cm⁻¹ v O-CH₃revealed that complexation has taken place and that azomethine and oxygen the of methoxygroups involved were in coordination to the metal ion. implied that the ligand L, was tridented.

The antimicrobial analysis of the compounds obtained (Tables 7a and 7b), revealed that the metal complexes have higher antibacterial activity than the free ligand. This suggests that chelation increases the antibacterial activity of the compounds as reported by [17, 18, 19].

CONCLUSION

(E)-4-methoxy-N-(4-methoxybenzylidene) aniline and its transition metal complexes have been synthesized in 2:1 molar ratio of ligand to metal ion. The complexes are all air stable and are anhydrous. Infrared studies have shown that L coordinated as a tridentate ligand. Job's method of mole ratio determination for the complexes revealed the stoichiometric compositions

for the complexes as 1:2 molar ratio (metal:ligand). Magnetic susceptibility studies showed the metals to be paramagnetic in nature for all the complexes. Antibacterial studies have indicated thatthe complexes showed higher activities than the free ligand. The results of these investigations support the

suggested structure of the metal complexes.

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